

Microbiology Laboratory Syllabus Winter 2018

Instructors	e-Mail	Days	Time	Section
ROOM 200 Labs				
Bianca Pereira	bianca.pereira@wayne.edu	T/Th	9:00 - 11:20 am	21602-003
Qing Chen	qing.chen2@wayne.edu	T/Th	11:30 am - 1:50 pm	21603-004
Duong Nguyen	dy1074@wayne.edu	T/Th	2:00 - 4:20 pm	21605-006
Mahmoud Suliman	fx2967@wayne.edu	T/Th	4:30 - 6:50 pm	21615-008
Abu Ramim	gg3939@wayne.edu	W/F	8:00 - 10:20 am	21601-002
Shashwat Mishra	fc2427@wayne.edu	W/F	11:30 am - 1:50 pm	21604-502
Tha Luong	thaluong.bio@gmail.com	W/F	2:00 - 4:20 pm	21606-007
Behdokht Jan Fada	ga2861@wayne.edu	W/F	4:30 - 6:50pm	21616-009
ROOM 404 Labs				
George Darany	gd5012@wayne.edu	T/Th	11:30 am - 1:50 pm	21619-013
Justin Dykema	gb5417@wayne.edu	T/Th	2:00 - 4:20 pm	21620-014
Becky Jackson	au7371@wayne.edu	W/F	8:00 - 10:20 am	22541-015
Becky Jackson	au7371@wayne.edu	W/F	11:30 am - 1:50 pm	21617-011
Sudeshna Biswas	gi1698@wayne.edu	W/F	2:00 - 4:20 pm	21618-012

Lab Location: RM 200 and RM 404 Shapero Hall

Course Objectives:

Upon the successful completion of this laboratory, the student should be able to:

1. Safely handle and grow pure cultures of microorganisms.
2. Perform standard microbial procedures such as dilutions, pipetting, and enumeration of microbial populations.
3. Use the light microscope correctly.
4. Isolate and identify microbes from natural samples such as soil and the human body.
5. Perform and interpret basic immunological and biochemical assays.

Lab Manual:

Microbiology: Laboratory Theory and Application 4th Edition Customized for Wayne State University

Michael J. Leboffe and Burton E. Pierce

ISBN-10: 1-61731-455-2 ISBN-13: 978-1-61731-455-1

Handouts:

In addition to the laboratory manual, some supplemental material will be provided as handouts for downloading off of the Lecture Section of the Blackboard website under General Lab Materials.

Grading:

Daily Quizzes. We will be using the i>Clicker system. **Students must purchase the i>clicker2 which is available at the Bookstore.** Students will be asked to answer 5 questions at the beginning of each lab. **Once the quiz starts, the doors to the lab will be closed, and students will not be permitted to enter the lab until after the quiz is finished.** Questions will cover the lab material assigned for that day and at minimum the lab day prior. The five lowest daily quiz scores will be dropped. Throughout the semester notebook prelab checks will be incorporated and will constitute the 5 points for that day's quiz. **Prelab checks that replace the daily quiz cannot be dropped.** During the first two weeks of the semester (while students are purchasing and registering their clickers), the Lab Instructors will be able to hand grade the questions for scoring. **Starting with the third week of lab, students must have and use their clickers to have their quiz answers scored. Quizzes will not be hand graded for any reason after this time.** Students must be sure to have their clickers registered and work out any user issues during the first two weeks of the semester while hand grading is available. Any students having issues with their clickers should notify their TA of the issue and should contact iClicker2 to have them resolved. Students with clicker issues after the second full week of classes will have to use any quizzes that can not be scored using the i>clicker2 as part of the five they are allowed to drop. In the event that a student

changes clickers during the semester, it is the student's responsibility to inform his/her TA that the clicker has been changed. Failure to inform TAs regarding clicker changes will result in the loss of points for the daily quizzes. At the end of the semester, the percentage of correct responses will be calculated and added to the final grade.

There are no makeups for quiz questions missed either because a student is late, absent, or because of user issues with the clickers.

Any student that is marked absent will receive a zero for their daily quiz score.

Laboratory notebooks. Keeping a well-organized and thorough laboratory notebook is an essential part of laboratory research. It is absolutely essential that you write your pre-lab: introduction, purpose statement, and protocols in your laboratory notebooks BEFORE you arrive at lab to do the actual experiments. Throughout the semester laboratory instructors will perform random pre-lab notebook inspections mentioned above to be sure that students are prepared to safely perform the laboratory experiments. Individuals who have not written their protocols in their notebooks will be penalized 5 points. Some of the experiments performed in lab will be completed in groups. It is the student's responsibility to obtain all results from all groups in the lab and incorporate these results in their notebooks. After the data from the experiments are obtained, the results, discussions and conclusions can be written to complete that entry in the notebook. Notebooks will also be collected and graded three times throughout the semester. **Notebooks will only be accepted in the laboratory classroom during your regular laboratory session.** Students will be notified one week prior to the date that notebooks are to be collected. Failure to turn in the notebook **at the beginning of lab** and during normal laboratory hours on the due date will result in a 10% penalty. An additional 10% penalty will be assessed **for each business day the notebooks are late.** The first time notebooks are collected the maximum grade will be 15 points. Each of the remaining two times the notebooks are collected the maximum grade will be 30 points. These grades will be based upon the criteria discussed in the *Keeping a Laboratory Notebook* handout. Students should have their notebooks with them each laboratory period with all sections except "results" and "discussion" completed before lab begins.

Laboratory Exams: Two major laboratory exams will be given. Students will be tested on the degree to which they have achieved the course objectives described above. The first exam will be given at mid-term and will be a combination of written (short answer and multiple choice) questions and some practical stations. The second exam will be entirely in the form of a laboratory practical and will be given during the last laboratory period. The laboratory practical will be comprehensive. These exams will include application of the skills and knowledge acquired during the semester to solve microbiological problems as well as evaluation of the student's technical competency and knowledge. Improperly or incompletely spelled names of microorganisms will be marked off.

No make-up exams will be given for laboratory midterms or practicals.

Electronic devices, other than an iClicker2, are not permitted in lab. Having such a device at your student bench during a daily quiz, the Midterm Exam, or the Final Lab Practical will be considered cheating, and the student will receive a failing grade for the course.

Grade Disputes:

A one (1) week period will be granted after the return of a notebook or lab exam, or the posting of your daily quiz score online to challenge a score. After the 1-week period, there will be no room to challenge your score. Students should submit any score challenges in writing to their laboratory instructor before this one-week period ends.

PLEASE NOTE: This week to challenge a score starts after the Graduate Teaching Assistant hands back the graded item or posts the quiz score online, not from the time at which you decide to pick it up or check your score on Blackboard.

It is the student's responsibility to check his/her scores on Blackboard. If a score is missing or incorrect, then the student must contact his/her TA in a timely manner. Scores will not be adjusted after the week to challenge has passed.

For the Final Lab Practical and final notebook collection, the time allotted for grade dispute will be adjusted to approximately two days after these items are handed back to allow for assigning of final grades by the Instructor.

Attendance:

Students are only permitted in the lab room while their lab instructor is present. Students should not enter the lab until given entry by their instructor.

Students can only attend the lab section **for which they are officially registered. Attendance in the laboratory portion of the course is mandatory.** Students who are absent from **one-fourth (7 or more)** laboratory periods, *total*, will not be allowed to continue in the laboratory and will automatically receive a grade of "0" for the laboratory portion of the course. Once a student has accrued the maximum number of absences, he or she will no longer be permitted to participate in the lab as it would be considered a safety hazard for the rest of the students in the course.

Students must contact their laboratory instructor via email as soon as possible in the event of their absence.

Students who leave the laboratory before completing their experiments or who come to lab more than 20 minutes late will be marked absent. Laboratory experiments that are missed due to a student's absence cannot be made up. Students who miss a lab should get the results from persons in their laboratory section.

In addition, students who are absent from **two or more consecutive** laboratory periods must obtain written permission from Dr. Higgs, in person during her office hours, before returning to class. This policy is designed to protect students and instructors from individuals who are not properly trained in the safe handling of microbes and laboratory equipment.

Students must wear their Personal Protection Equipment (lab coats and goggles) and have proper attire when in lab. If you do not have your lab coat or you are not wearing proper attire, then you will not be allowed into the lab. You will also be marked absent and will not be allowed to answer the daily quiz questions or hand in your notebook. Please refer to the Lab Safety Rules handout for more information regarding proper dress code while in lab.

Any student that is asked to leave the lab, for any reason, will be marked absent and be given a zero for their daily quiz score.

Failure to wear proper attire on the days of exams will, at minimum, result in a penalty of 10% of the total points possible on the exam.

Microbiology Laboratory Schedule

Lab Book

Week	Lab	Date	Experiment/Exercise	Experiment	Pages
1	1	T. January 9 W. January 10	Check-in Lab safety, syllabus and course requirements. Reading assignment - read and complete Microscopy exercise posted on Blackboard	Handouts and Intro Handout	1-9
1	2	Th. January 11 F. January 12	Microscopy and Survey of Microorganisms Assign microscopes Ubiquity of Microorganisms; part I Introduction to the Light Microscope	2-1 3-1 (no pre-lab needed) Microscopy Exercise replaces pre-lab	55,57-58 141-149
LAB COATS REQUIRED BEFORE NEXT LABORATORY					
2	3	T. January 16 W. January 17	Microscope Slide Techniques; Bacterial Morphology <i>Turn in completed Microscopy Handout before class begins.</i> Common Aseptic Transfers and Inoculation Methods Ubiquity of Microorganisms: Cultural Characteristics; part II Bacterial Structure and Simple Staining	Microscopy Handout 1-3 (Read Only) -- no prelab needed 2-2, 2-4 (use terms from these exercises in the results section of Ex. 2-1) 3-5	27; 29-38 63-72; 79 181-188
2	4	Th. January 18 F. January 19	Bacterial Morphology and Cytology Negative Stain Capsular Staining	3-6 3-9	191-192 211-212
Starting with the next laboratory session, Daily quiz questions will be scored via the i-clickers. Students must purchase and register their i-clickers in order to have their answers scored.					
3	5	T. January 23 W. January 24	Bacterial Morphology and Cytology The Gram Stain Motility Test: semisolid media	3-7 5-28	195-200 437-438
3	6	Th. January 25 F. January 26	Record results of Exercise 5-28 Bacterial Morphology and Cytology Acid-Fast Stain (Ziehl-Neelsen Method) Bacterial Endospores (Schaeffer-Fulton Method)	5-28 3-8 3-10	437-438 203-207 215-218
4	7	T. January 30 W. January 31	Culture Methods: Preparation of microbial media. Culture Media Preparation and Steam Sterilization.	1-2	21-26
4	8	Th. February 1 F. February 2	Microbial Nutrition and Pure Culture Methods Microbial Growth Requirements Streak plate (omit pour plate technique) Selective and Differential Media Phenylethyl Alcohol Agar Mannitol Salt Agar MacConkey Agar Eosin Methylene Blue Agar Hektoen Enteric Agar Biosafety Cabinet	1-2 1-4 4-1 4-4 4-5 4-6 4-7 Refer to Lab Lecture	21-26 41-46 235-238 253-254 259-262 267-268 273-274
5	9	T. February 6 W. February 7	Microbial Nutrition and Pure Culture Methods <i>Record results of streak plate and where growth is present</i> Pick isolated colonies from streak plate and make pure cultures. Selective and Differential Media <i>Record results of Ex. 4-1, Ex. 4-4, Ex. 4-5, Ex. 4-6, Ex. 4-7</i> Eukaryotic Microbes: The Fungi The Fungi- Common Yeasts and Molds (Day #1)	1-4 1-4 4-1, 4-4, 4-5, 4-6, 4-7 12-1	25-28 25-28 235-238; 253-254; 259-262; 267-268; 273-274 783-791
5	10	Th. February 8 F. February 9	Check pure cultures for good isolation and record results. Eukaryotic Microbes: The Fungi The Fungi- Common Yeasts and Molds (Day #2) Bacterial Population Counts Standard Plate Count (Viable Count) Using Glass Pipettes Bacterial Viruses Viruses: Isolation of Bacteriophage from Sewage: Enrichment	12-1 12-1 6-1 Appendix C (read only) 6-5	783-791 467-472 839-842 497-501
6	11	T. February 13 W. February 14	Results of Standard Plate Count (Viable Count) Bacterial Viruses Viruses: Isolation of Bacteriophage from Sewage: Filtration and Seeding	6-1 6-5	467-472 497-501
6	12	Th. February 15 F. February 16	Mid-term Written Examination: 75 points Mid-term Written Examination: 75 points		

Week	Lab	Date	Experiment/Exercise	Lab Book	
				Experiment	Pages
7	13	T. February 20 W. February 21	Bacterial Viruses <i>Determine Titer of virus in samples</i> <i>Determine the efficacy of membrane filtration</i> Environmental Influences on Microbial Growth Oxygen requirements of microorganisms Temperature: Effects on Growth Osmotic Pressure and Bacterial Growth Identification of Bacteria: Unknowns Day #1	6-5 6-5 2-6, 2-7,2-8 2-9 2-11 Refer to the lab lecture	497-501 497-501 91-92,95-96,99-100 103-104 113-115
7	14	Th. February 22 F. February 23	<i>Record results of Exercises 2-6, 2-7,2-8,2-9</i> <i>Record results of Exercise 2-9</i> <i>Record results of Exercise 2-11</i> Identification of Bacteria: Unknowns Day #2 Morphology and Cultural Characteristics	2-6, 2-7,2-8 2-9 2-11 Refer to the lab lecture	91-92,95-96,99-100 103-104 113-115
8	15	T. February 27 W. February 28	Identification of Bacteria <i>Record results of Unknowns Day#2</i> Identification of Bacteria: Unknowns Day #3 Physiological Characteristics: Oxidation and Fermentation Tests	Refer to the lab lecture 5-3, 5-4, 5-6, 5-8	303-306,311-314,321-323, 333-336
8	16	Th. March 1 F. March 2	Identification of Bacteria Record results of Unknowns Day #3 Physiological Characteristics: Miscellaneous Tests (Day #4)	Refer to the lab lecture 5-9, 5-12, 5-20, 5-21	339-341, 357-358, 393-396, 401-403
9	17	T. March 6 W. March 7	Identification of Bacteria <i>Record results of Unknowns Day#4</i> Identification of Bacteria: Unknowns Day #5 Physiological Characteristics: Hydrolytic Reactions	5-13, 5-15, 5-16, 5-18	361-363, 371-372, 375-376, 383-385
9	18	Th. March 8 F. March 9	<i>Record results of Unknowns Day#5</i> Form a Hypothesis regarding the identify of your Unknown bacterium The API 20E Identification System	Refer to the lab lecture 5-29	441-446
10	19	T. March 13 W. March 14	No Classes -- Spring Break No Classes -- Spring Break		
10	20	Th. March 15 F. March 16	No Classes -- Spring Break No Classes -- Spring Break		
11	21	T. March 20 W. March 21	<i>Add reagents and record results of API-20E Test Strip</i> <i>Identify unknown based upon API Analytical Profile Index</i> Molecular Diagnostics and Cloning: Practice with Pipettors Transfers using a digital pipette	5-29 5-29 Refer to the lab lecture Appendix D (read only)	441-446 441-446 843-846 (read only)
11	22	Th. March 22 F. March 23	Molecular Diagnostics and Cloning: PCR	10-4	713-720
12	23	T. March 27 W. March 28	Agarose gel electrophoresis and restriction digestion Confirm PCR product -- Agarose Gel Electrophoresis Digest PCR product and cloning vector with restriction endonuclease	Refer to the lab lecture Appendix G (read only) 10-2	857-862 693-698
12	24	Th. March 29 F. March 30	Gel analysis of restriction digests and DNA Analyze digestion pattern of digested PCR product: Identify microorganism Ligate PCR products to digested vector	Refer to the lab lecture	
13	25	T. April 3 W. April 4	Transform E. coli with ligated DNA Transform E. coli DH5 cells with ligated DNA and transform E. coli Plate transformed cells on selective medium	Refer to the lab lecture	
13	26	Th. April 5 F. April 6	Replicate transformants to identify vectors containing cloned DNA Ultraviolet Light: Lethal Effects Antibiotic Sensitivity Testing	Refer to the lab lecture 2-13 7-3	64-65 529-533
14	27	T. April 10 W. April 11	<i>Record results of Exercise 2-13</i> <i>Record results of Exercise 7-3</i> Medical Microbiology and Immunology Identification of Gram Positive Pathogens: Staphylococcus Day#1 E-Test and MICs Day #1	2-13 7-3 Refer to the lab lecture 4-4, 5-14, 5-25, 5-27 Refer to the lab lecture	64-65 529-533 253-254, 367-368, 423-425, 433-434
14	28	Th. April 12 F. April 13	Medical Microbiology and Immunology Read Results of Identification of Gram Positive Pathogens Identification of Gram Positive Pathogens: Staphylococcus Day#2 E-Test and MICs Day #2	Refer to the lab lecture Refer to the lab lecture Refer to the lab lecture	
15	29	T. April 17 W. April 18	Medical Microbiology and Immunology Record Results of E-test and MICs Review for Practical		
15	30	Th. April 19 F. April 20	LABORATORY FINAL - LAB PRACTICAL: Comprehensive, 150 points LABORATORY FINAL - LAB PRACTICAL: Comprehensive, 150 points		