Microbiology Laboratory Syllabus Fall 2019

Instructors	e-Mail	Days	Time	Section
ROOM 200 Labs				
Dave Lall	DaveLall@wayne.edu	T/Th	8:30 - 10:50 am	12123-006
Abu Ramim	gg3939@wayne.edu	T/Th	11:00 am-1:20 pm	12124-007/12139-014
Sudeshna Biswas	gi1698@wayne.edu	T/Th	1:30 - 3:50 pm	12125-008/12140-016
Qing Chen	ej1656@wayne.edu	W/F	8:30 - 10:50 am	12119-002
Imjoo Jang	ep8358@wayne.edu	W/F	11:00 am-1:20 pm	12120-502
Chisom Joshua Onu	chisomjoshua@wayne.edu	W/F	2:30 - 4:50 pm	12121-004
ROOM 404 Labs				
Muntasir Rahman	gp6161@wayne.edu	W/F	11:00 am - 1:20 pm	12128-011
Linh Vo	gn4785@wayne.edu	W/F	2:30 - 4:50 pm	16279-012

Lab Location: RM 200 and RM 404 Shapero Hall

Course Objectives:

Upon the successful completion of this laboratory, the student should be able to:

- 1. Safely handle and grow pure cultures of microorganisms.
- 2. Perform standard microbial procedures such as dilutions, pipetting, and enumeration of microbial populations.
- 3. Use the light microscope correctly.
- 4. Isolate and identify microbes from natural samples such as soil and the human body.
- 5. Perform and interpret basic immunological and biochemical assays.

Lab Manual:

Microbiology: Laboratory Theory and Application 4th Edition Customized for Wayne State University

Michael J. Leboffe and Burton E. Pierce

ISBN-10: 1-61731-455-2 ISBN-13: 978-1-61731-455-1

Handouts:

In addition to the laboratory manual, some supplemental material will be provided as handouts for downloading off of your laboratory section of Canvas.

Grading:

Daily Quizzes. We will be using the i>Clicker system. Students must purchase the i>clicker2, which is available at the Bookstore. Students will be asked to answer 5 questions at the beginning of each lab. Once the quiz starts, the doors to the lab will be closed, and students will not be permitted to enter the lab until after the quiz is finished. Questions will cover the lab material assigned for that day and at minimum the lab day prior. The five lowest daily quiz scores will be dropped.

During the first two weeks of the semester (while students are purchasing and registering their clickers), the Lab Instructors will be able to hand grade the questions for scoring. Starting with the third week of lab, students must have and use their clickers to have their quiz answers scored. Quizzes will not be hand graded for any reason after this time. Students must be sure to have their clickers registered and work out any user issues during the first two weeks of the semester while hand grading is available. Any students having issues with their clickers should notify their TA of the issue and should contact iClicker2 to have them resolved. Students with clicker issues after the second full week of classes will have to use any quizzes that can not be scored using the i>clicker2 as part of the five they are allowed to drop. In the event that a student changes clickers during the semester, it is the student's responsibility to inform his/her TA that the clicker has been changed. Failure to inform TAs regarding clicker changes will result in the loss of points for the daily quizzes. At the end of the semester, the percentage of correct responses will be calculated and added to the final grade.

There are no makeups for quiz questions missed either because a student is late, absent, or because of user issues with the clickers.

Any student that is marked absent will receive a zero for their daily quiz score.

<u>Laboratory notebooks.</u> Keeping a well-organized and thorough laboratory notebook is an essential part of laboratory research. It is absolutely essential that you write your pre-lab: introduction, purpose statement, and protocols in your laboratory notebooks BEFORE you arrive at lab to do the actual experiments. Some of the experiments performed in lab will be completed in groups. It is the student's responsibility to obtain all results from all groups in the lab and incorporate these results in their notebooks. After the data from the experiments are obtained, the results, discussions and conclusions can be written to complete that entry in the notebook.

<u>Pre-lab write-up checks</u>. Pre-lab write-ups are worth 25 points of your total laboratory score. Throughout the semester laboratory instructors will perform random pre-lab write-up inspections to be sure that students are prepared to safely perform the laboratory experiments. Each pre-lab check is worth up to 5 points.

Notebooks will only be accepted in the laboratory classroom during your regular laboratory session. Students will be notified one week prior to the date that notebooks are to be collected. Failure to turn in the notebook at the beginning of lab and during normal laboratory hours on the due date will result in a 10% penalty. An additional 10% penalty will be assessed for each business day the notebooks are late. The first time notebooks are collected the maximum grade will be 25 points. Each of the remaining two times the notebooks are collected the maximum grade will be 35 points. These grades will be based upon the criteria discussed in the *Keeping a Laboratory Notebook* handout. Students should have their notebooks with them each laboratory period with all sections except "results" and "discussion" completed before lab begins.

<u>Laboratory Exams</u>: Two major laboratory exams will be given. Students will be tested on the degree to which they have achieved the course objectives described above. The first exam will be given at midterm and will be a combination of written (short answer and multiple choice) questions and some practical stations. The second exam will be entirely in the form of a laboratory practical and will be given during the last laboratory period. The laboratory practical will be comprehensive. These exams will include application of the skills and knowledge acquired during the semester to solve microbiological problems as well as evaluation of the student's technical competency and knowledge. Improperly or incompletely spelled names of microorganisms will be marked off.

No make-up exams will be given for laboratory midterms or practicals.

Electronic devices, other than an iClicker2, are not permitted in lab. Having such a device at your student bench during a daily quiz, the Midterm Exam, or the Final Lab Practical will be considered cheating, and the student will receive a failing grade for the course.

Students are not permitted to have anything on their wrist or in their ears during quizzes, exams, or practicals. All watches, wrist jewelry, ear buds, etc. must be removed prior to the start of the quiz, exam, or practical.

Students must wear appropriate footwear and apparel on test days. Any student who comes to lab inappropriately dressed will face a 10% deduction from their exam score.

Grade Disputes:

A one (1) week period will be granted after the return of a notebook or lab exam, or the posting of your daily quiz score online to challenge a score. After the 1-week period, there will be no room to challenge your score. Students should submit any score challenges in writing to their laboratory instructor before this one-week period ends.

PLEASE NOTE: This week to challenge a score starts after the Graduate Teaching Assistant hands back the graded item or posts the quiz score online, not from the time at which you decide to pick it up or check your score on Canvas.

It is the student's responsibility to check his/her scores on Canvas. If a score is missing or incorrect, then the student must contact his/her TA in a timely manner. Scores will not be adjusted after the week to challenge has passed.

For the Final Lab Practical and final notebook collection, the time allotted for grade dispute will be adjusted to approximately two days after these items are handed back to allow for assigning of final grades by the Instructor.

Attendance:

Students are only permitted in the lab room while their lab instructor is present. Students should not enter the lab until given entry by their instructor.

Students can only attend the lab section <u>for which they are officially registered</u>. <u>Attendance in the laboratory portion of the course is mandatory</u>. Students who are absent from <u>one-fourth (7 or more)</u> laboratory periods, *total*, will not be allowed to continue in the laboratory and will automatically receive a grade of "0" for the laboratory portion of the course. Once a student has accrued the maximum number of absences, he or she will no longer be permitted to participate in the lab as it would be considered a safety hazard for the rest of the students in the course.

<u>Please note</u>: We will not make any exceptions to this attendance policy, so please use your absences wisely.

Students must contact their laboratory instructor via email as soon as possible in the event of their absence.

Students who leave the laboratory before completing their experiments or who come to lab more than 20 minutes late will be marked absent. Laboratory experiments that are missed due to a student's absence cannot be made up. Students who miss a lab should get the results from persons in their laboratory section.

In addition, students who are absent from **two or more** *consecutive* laboratory periods must obtain written permission from Dr. Schrader, in person during his office hours, before returning to class. This policy is designed to protect students and instructors from individuals who are not properly trained in the safe handling of microbes and laboratory equipment.

Students must wear their Personal Protection Equipment (lab coats and goggles) and have proper attire when in lab. If you do not have your lab coat or you are not wearing proper attire, then you will not be allowed into the lab. You will also be marked absent and will not be allowed to answer the daily quiz questions or hand in your notebook. Please refer to the Lab Safety Rules handout for more information regarding proper dress code while in lab.

Any student that is asked to leave the lab, for any reason, will be marked absent and be given a zero for their daily quiz score.

Failure to wear proper attire on the days of exams will, at minimum, result in a penalty of 10% of the total points possible on the exam.

<u>Lab Issues:</u> All lab concerns should first be brought to the attention of your lab instructor, then the lab coordinator (Krystyn Purvis), and then the lecturer for the course.

Krystyn Purvis' email address: Krystyn@wayne.edu

Lab Rules:

Students are responsible for reading and abiding by the rules stated in the Laboratory Safety Rules handout. Failure to follow these rules could result in loss of points, removal from the lab, or a failing grade in the course.

Summary of Laboratory Grading Criteria

Pre-lab check 25 points
Microscopy Ex. 5 points
Reading Assignments 25 points

Lab Notebooks 95 points (25+35+35)

Midtern Exam100 pointsFinal Lab Practical150 pointsLab Total400 points

Laboratory Supplies:

Research Laboratory Notebook (see above)

Lab Coat (Laboratory Safety Rules)

Blue, Sharpie marker (other colors are not permitted) iClicker 2 Splash-proof lab goggles (Students are not permitted to use safety glasses.)

All other supplies will be provided.

Your laboratory instructor will provide information regarding the purchase of lab coats and notebooks.

Students with disabilities:

If you have a documented disability that requires accommodation, please register with Student Disability Services for coordination of your academic accommodations. The Student Disability Services (SDS) office is located at 1600 David Adamany Undergraduate Library within the Student Academic Success Services Department.

The SDS telephone number is 313-577-1851 or 313-577-3365; (TTY--telecommunication device for hearing impaired students).

Once you have your accommodations in place, I will be happy to meet with you privately during office hours to discuss your needs.

Please be aware that a delay in getting this documentation may also delay the facilitation of your needed accommodations. Accommodations and services cannot be guaranteed if students choose not to follow the procedures for registering with Student Disability Services in a timely

manner. Accommodations and services can be revisited as needed, but they <u>are not retroactive</u> and cannot be guaranteed if procedures are not followed with reasonable, advanced notice.

Student Disability Services' mission is to assist the university in creating an accessible community where students with disabilities have an equal opportunity to fully participate in their educational experience at Wayne State University.

Please refer to the SDS website for further information about students with disabilities and the services provided for faculty and students: http://studentdisability.wayne.edu/

Students with accommodation letters must provide their instructor and the lab coordinator with the letter prior to the third week of lab.

Microbiology Laboratory Schedule

1 1	Th. August 29 F. August 30 B COATS REQUIRED BEFORE NEX T. September 3 W. September 4 Th. September 5 F. September 6 Starting with the next Students must purch T. September 10 W. September 11 Th. September 12 F. September 13	LAB COA	Experiment/Exercise Check-in Lab safety, syllabus and course requirements. Reading assignment - read and complete Microscopy exercise posted on Blackboard ABORATORY Microscopy and Survey of Microorganisms Assign microscopes Ubiquity of Microorganisms; part I Introduction to the Light Microscope Microscopy Exercise replaces pre-lab Microscope Slide Techniques; Bacterial Morphology Turn in completed Microscopy Handout before class begins. Common Aseptic Transfers and Inoculation Methods Microscope Slide Techniques; Bacterial Morphology Turn in completed Microscopy Handout before class begins. Common Aseptic Transfers and Inoculation Methods Ubiquity of Microorganisms: Cultural Characteristics; part II Bacterial Structure and Simple Staining Oratory session, Daily quiz questions will be scored via the i-clickers. and register their i-clickers in order to have their answers scored.	Pages 1-9 55,57-58 141-149 27; 29-38 63-72; 79 181-188
LAB 2 2 2 2 3 3 4 4 3 5 4 6 4 7 5 8	F. August 30 B COATS REQUIRED BEFORE NEX T. September 3 W. September 4 Th. September 5 F. September 6 Starting with the next Students must purch: T. September 10 W. September 11 Th. September 12 F. September 13	LAB COA	Lab safety, syllabus and course requirements. Reading assignment - read and complete Microscopy exercise posted on Blackboard ABORATORY Microscopy and Survey of Microorganisms Assign microscopes Ubiquity of Microorganisms; part I Introduction to the Light Microscope Microscopy Exercise replaces pre-lab Microscope Slide Techniques; Bacterial Morphology Turn in completed Microscopy Handout before class begins. Common Aseptic Transfers and Inoculation Methods Microscope Slide Techniques; Bacterial Morphology Turn in completed Microscopy Handout before class begins. Common Aseptic Transfers and Inoculation Methods Ubiquity of Microorganisms: Cultural Characteristics; part II Bacterial Structure and Simple Staining Diratory session, Daily quiz questions will be scored via the i-clickers.	55,57-58 141-149 27; 29-38 63-72; 79
2 2 2 2 3 3 4 3 5 4 6 4 7 5 8	T. September 3 W. September 4 Th. September 5 F. September 6 Starting with the next Students must purch T. September 10 W. September 11 Th. September 12 F. September 13	3	Microscopy and Survey of Microorganisms Assign microscopes Ubiquity of Microorganisms; part I Introduction to the Light Microscope Microscope Slide Techniques; Bacterial Morphology Turn in completed Microscopy Handout before class begins. Common Aseptic Transfers and Inoculation Methods Common Aseptic Transfers and Inoculation Methods Ubiquity of Microorganisms: Cultural Characteristics; part II Bacterial Structure and Simple Staining Diratory session, Daily quiz questions will be scored via the i-clickers.	141-149 27; 29-38 63-72; 79
2 3 3 4 3 5 4 6 4 7 5 8	W. September 4 Th. September 5 F. September 6 Starting with the next Students must purch: T. September 10 W. September 11 Th. September 12 F. September 13	3	Assign microscopes Ubiquity of Microorganisms; part I Introduction to the Light Microscope Microscope Slide Techniques; Bacterial Morphology Turn in completed Microscopy Handout before class begins. Common Aseptic Transfers and Inoculation Methods Ubiquity of Microorganisms: Cultural Characteristics; part II Bacterial Structure and Simple Staining Dratory session, Daily quiz questions will be scored via the i-clickers.	141-149 27; 29-38 63-72; 79
3 4 3 5 4 6 4 7 5 8	Th. September 5 F. September 6 Starting with the next Students must purch T. September 10 W. September 11 Th. September 12 F. September 13		Ubiquity of Microorganisms; part I Introduction to the Light Microscope Microscope Slide Techniques; Bacterial Morphology Turn in completed Microscopy Handout before class begins. Common Aseptic Transfers and Inoculation Methods Common Aseptic Transfers and Inoculation Methods Ubiquity of Microorganisms: Cultural Characteristics; part II Bacterial Structure and Simple Staining Dratory session, Daily quiz questions will be scored via the i-clickers.	141-149 27; 29-38 63-72; 79
3 4 3 5 4 6 4 7 5 8	Starting with the next Students must purch. T. September 10 W. September 11 Th. September 12 F. September 13		Introduction to the Light Microscope 3-1 (no pre-lab needed) Microscope Slide Techniques; Bacterial Morphology Turn in completed Microscopy Handout before class begins. Common Aseptic Transfers and Inoculation Methods 1-3 (Read Only) no prelab needed 2-2, 2-4 (use terms from these exercises in the results section Ubiquity of Microorganisms: Cultural Characteristics; part II Bacterial Structure and Simple Staining 3-5 Diratory session, Daily quiz questions will be scored via the i-clickers.	141-149 27; 29-38 63-72; 79
3 4 3 5 4 6 4 7 5 8	Starting with the next Students must purch. T. September 10 W. September 11 Th. September 12 F. September 13		Microscopy Exercise replaces pre-lab Microscopy Slide Techniques; Bacterial Morphology Turn in completed Microscopy Handout before class begins. Common Aseptic Transfers and Inoculation Methods Common Aseptic Transfers and Inoculation Methods Ubiquity of Microorganisms: Cultural Characteristics; part II Bacterial Structure and Simple Staining Oratory session, Daily quiz questions will be scored via the i-clickers.	27; 29-38 63-72; 79
3 4 3 5 4 6 4 7 5 8	Starting with the next Students must purch. T. September 10 W. September 11 Th. September 12 F. September 13		Microscope Slide Techniques; Bacterial Morphology Turn in completed Microscopy Handout before class begins. Common Aseptic Transfers and Inoculation Methods 1-3 (Read Only) no prelab needed 2-2, 2-4 (use terms from these exercises in the results section Ubiquity of Microorganisms: Cultural Characteristics; part II Bacterial Structure and Simple Staining 3-5 pratory session, Daily quiz questions will be scored via the i-clickers.	63-72; 79
3 4 3 5 4 6 4 7 5 8	Starting with the next Students must purch. T. September 10 W. September 11 Th. September 12 F. September 13		Turn in completed Microscopy Handout before class begins. Common Aseptic Transfers and Inoculation Methods 1-3 (Read Only) no prelab needed 2-2, 2-4 (use terms from these exercises in the results section Ubiquity of Microorganisms: Cultural Characteristics; part II Bacterial Structure and Simple Staining oratory session, Daily quiz questions will be scored via the i-clickers.	63-72; 79
3 4 3 5 4 6 4 7 5 8	Starting with the next Students must purch. T. September 10 W. September 11 Th. September 12 F. September 13		Turn in completed Microscopy Handout before class begins. Common Aseptic Transfers and Inoculation Methods 1-3 (Read Only) no prelab needed 2-2, 2-4 (use terms from these exercises in the results section Ubiquity of Microorganisms: Cultural Characteristics; part II Bacterial Structure and Simple Staining oratory session, Daily quiz questions will be scored via the i-clickers.	63-72; 79
3 5 4 6 4 7 5 8	Starting with the next Students must purch: T. September 10 W. September 11 Th. September 12 F. September 13	4	Common Aseptic Transfers and Inoculation Methods 1-3 (Read Only) no prelab needed 2-2, 2-4 (use terms from these exercises in the results section Ubiquity of Microorganisms: Cultural Characteristics; part II Bacterial Structure and Simple Staining 3-5 pratory session, Daily quiz questions will be scored via the i-clickers.	63-72; 79
3 5 4 6 4 7 5 8	T. September 10 W. September 11 Th. September 12 F. September 13	4	2-2, 2-4 (use terms from these exercises in the results section Ubiquity of Microorganisms: Cultural Characteristics; part II of Ex. 2-1) Bacterial Structure and Simple Staining 3-5 pratory session, Daily quiz questions will be scored via the i-clickers.	63-72; 79
3 5 4 6 4 7 5 8	T. September 10 W. September 11 Th. September 12 F. September 13	4	Ubiquity of Microorganisms: Cultural Characteristics; part II of Ex. 2-1) Bacterial Structure and Simple Staining 3-5 pratory session, Daily quiz questions will be scored via the i-clickers.	
3 5 4 6 4 7 5 8	T. September 10 W. September 11 Th. September 12 F. September 13	4	Ubiquity of Microorganisms: Cultural Characteristics; part II of Ex. 2-1) Bacterial Structure and Simple Staining 3-5 pratory session, Daily quiz questions will be scored via the i-clickers.	
3 5 4 6 4 7 5 8	T. September 10 W. September 11 Th. September 12 F. September 13	4	Bacterial Structure and Simple Staining 3-5 paratory session, Daily quiz questions will be scored via the i-clickers.	
3 5 4 6 4 7 5 8	T. September 10 W. September 11 Th. September 12 F. September 13	4		
3 5 4 6 4 7 5 8	W. September 11 Th. September 12 F. September 13	4		
3 5 4 6 4 7 5 8	W. September 11 Th. September 12 F. September 13		Bacterial Morphology and Cytology	
4 6 4 7 5 8	Th. September 12 F. September 13		Negative Stain 3-6	191-192
4 6 4 7 5 8	F. September 13		Capsular Staining 3-9	211-212
4 6 4 7 5 8	F. September 13			
4 7 5 8		5	Bacterial Morphology and Cytology	
4 7 5 8	T. September 17		The Gram Stain 3-7	195-200
4 7 5 8	T. September 17		Motility Test: semisolid media 5-28	437-438
4 7 5 8		6	Record results of Exercise 5-28 5-28	437-438
5 8	W. September 18		Bacterial Morphology and Cytology	
5 8			Acid-Fast Stain (Ziehl-Neelsen Method) 3-8	203-207
5 8			Bacterial Endospores (Schaeffer-Fulton Method) 3-10	215-218
5 8	Th. September 19	7	Culture Methods: Drengration of microbial media	
	F. September 20	,	Culture Methods: Preparation of microbial media. Culture Media Preparation and Steam Sterilization. 1-2	21-26
			Culture in Culture in Country	2.7.20
5 9		8	Microbial Nutrition and Pure Culture Methods	
5 9	W. September 25		Microbial Growth Requirements 1-2	21-26
5 9			Streak plate (omit pour plate technique) 1-4	41-46
5 9			Selective and Differential Media Phenylethyl Alcohol Agar 4-1	235-238
5 9			Mannitol Salt Agar 4-4	253-254
5 9			MacConkey Agar 4-5	259-262
5 9			Eosin Methylene Blue Agar 4-6	267-268
5 9			Hektoen Enteric Agar 4-7	273-274
5 9			Biosafety Cabinet Refer to Lab Lecture	
5 9				
	Th. September 26	9	Microbial Nutrition and Pure Culture Methods	
	F. September 27		Record results of streak plate and where growth is present 1-4	25-28
			Pick isolated colonies from streak plate and make pure cultures. 1-4	25-28
			Selective and Differential Media	
				235-238; 253-254; 259-26
			Record results of Ex. 4-1, Ex. 4-4, Ex. 4-5, Ex. 4-6, Ex. 4-7 4-1, 4-4, 4-5,4-6, 4-7	267-268; 273-274
			Eukaryotic Microbes: The Fungi	
			The Fungi- Common Yeasts and Molds (Day #1)	783-791
6 10	0 T. October 1	10	Check pure cultures for good isolation and record results.	
	W. October 2		Eukaryotic Microbes: The Fungi	
			TI 5 10 10 10 10 10 10 10 10 10 10 10 10 10	700 704
			The Fungi- Common Yeasts and Molds (Day #2) 12-1	783-791
		Bacterial Population Counts		
			Standard Plate Count (Viable Count) 6-1	467-472
			Using Glass Pipettes Appendix C (read only)	839-842
			Using Glass Pipettes Appendix C (read only) Bacterial Viruses	033-042
			Viruses: Isolation of Bacteriophage from Sewage: Enrichment 6-5	497-501
				137-301
6 11			Results of Standard Plate Count (Viable Count) 6-1	467-472
		11	Bacterial Viruses	
	1 Th. October 3 F. October 4	11	Viruses: Isolation of Bacteriophage from Sewage: Filtration and Seeding 6-5	497-501
7 12		11		
	F. October 4	11	Mid-term Written Examination: 75 points	

Week	Lab	Date	Experiment/Exercise	Experiment	Pages
7	13	Th. October 10 F. October 11	Bacterial Viruses Determine Titer of virus in samples	6-5	497-501
			Determine the efficacy of membrane filtration	6-5	497-501
			Environmental Influences on Microbial Growth Oxygen requirements of microorganisms	2-6, 2-7,2-8	91-92,95-96,99-100
			Temperature: Effects on Growth	2-9	103-104
			Osmotic Pressure and Bacterial Growth Identification of Bacteria: Unknowns Day #1	2-11 Refer to the lab lecture	113-115
			identification of bacteria. Officiowills bay #1	Refer to the lab lecture	
8	14	T. October 15	Record results of Exercises 2-6, 2-7,2-8,2-9	2-6, 2-7,2-8	91-92,95-96,99-100
		W. October 16	Record results of Exercise 2-9 Record results of Exercise 2-11	2-9 2-11	103-104 113-115
			Identification of Bacteria:		
			Unknowns Day #2 Morphology and Cultural Characteristics	Refer to the lab lecture	
8	15	Th. October 17	Identification of Bacteria		
-		F. October 18	Record results of Unknowns Day#2		
			Identification of Bacteria: Unknowns Day #3	Refer to the lab lecture	303-306,311-314,321-323,
			Physiological Characteristics: Oxidation and Fermentation Tests	5-3, 5-4, 5-6, 5-8	333-336
9	16	T. October 22	Identification of Bacteria		
		W. October 23	Record results of Unknowns Day #3	Refer to the lab lecture	339-341, 357-358, 393-396,
			Physiological Characteristics: Miscellaneous Tests (Day #4)	5-9, 5-12, 5-20, 5-21	401-403
9	17	Th. October 24	Identification of Bacteria		
		F. October 25	Record results of Unknowns Day#4		
			Identification of Bacteria: Unknowns Day #5		361-363, 3/1-3/2, 3/5-3/6,
			Physiological Characteristics: Hydrolytic Reactions	5-13, 5-15, 5-16. 5-18	383-385
			Form a Hypothesis regarding the identify of your Unknown bacterium		
			The API 20E Identification System	5-29	441-446
10	18	T. October 29	Record results of Unknowns Day#5	Refer to the lab lecture	
10	10	W. October 30	Add reagents and record results of API-20E Test Strip	5-29	441-446
			Identify unknown based upon API Analytical Profile Index Molecular Diagnostics and Cloning: Practice with Pipettors Tranfers using a digital pipette	5-29 Refer to the lab lecture	441-446
				Appendix D (read only)	843-846 (read only)
10	19	Th. October 31 F. November 1	Molecular Diagnostics and Cloning: PCR	10-4 Refer to the lab lecture	713-720
11	20	T. November 5 W. November 6	Agarose gel electrophoresis and restriction digestion	Refer to the lab lecture Appendix G (read only)	857-862
		w. November 6	Confirm PCR product Agarose Gel Electrophoresis Digest PCR product and cloning vector with restriction endonuclease	10-2	693-698
11 21	Th. November 7 F. November 8	Gel analysis of restriction digests and DNA Analyze digestion pattern of digested PCR product: Identify microorganism	Refer to the lab lecture		
		1. November 6	Ligate PCR products to digested vector		
12	22	T. November 12	Transfrom E. coli with ligated DNA	Refer to the lab lecture	
12	22	W. November 13	Transfrom E. coli with ligated DNA Transform E. coli DH5 cells with ligated DNA and transfrom E. coli	Refer to the lab lecture	
			Plate transformed cells on selective medium		
12	23	Th. November 14	Replicaplate transformants to identify vectors containing cloned DNA	Refer to the lab lecture	
		F. November 15	Ultraviolet Light: Lethal Effects	2-13	64-65
			Antibiotic Sensitivity Testing	7-3	529-533
13	24	T. November 19 W. November 20	Record results of Replica-plating	Refer to the lab lecture	64.65
	W. November 20	Record results of Exercise 2-13 Record results of Exercise 7-3	2-13 7-3	64-65 529-533	
			Medical Microbiology and Immunology Identification of Gram Positive Pathogens: Staphylococcus Day#1	Refer to the lab lecture	
			dentineation of drain out of a thoughts. Staphylococcus bay#1		253-254, 367-368, 423-425,
			E-Test and MICs Day #1	4-4, 5-14, 5-25, 5-27 Refer to the lab lecture	433-434
			- · · · · · · · · · · · · · · · · · · ·		
	25	71 11 1 04			
13	25	Th. November 21 F. November 22	Medical Microbiology and Immunology Read Results of Identification of Gram Positive Pathogens	Refer to the lab lecture	
			Identification of Gram Positive Pathogens: Staphylococcus Day#2 E-Test and MICs Day #2	Refer to the lab lecture	
			E-Test and Mics Day #2	Refer to the lab lecture	
14	26	T. November 26 W. November 27	No Labs No Labs Thanksgiving Holiday Closure		
			The last of the la		
14	27	Th. November 28	No Labs Thanksgiving Holiday Closure		
		F. November 29	No Labs Thanksgiving Holiday Closure		
15	28	T. December 3	Medical Microbiology and Immunology		
-	-	W. December 4	Record Results of E-test and MICs		
			Review for Practical		
15	29	Th. December 5 F. December 6	LABORATORY FINAL - LAB PRACTICAL: Comprehensive, 150 points LABORATORY FINAL - LAB PRACTICAL: Comprehensive, 150 points		
		December o	To		